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AWARD NUMBER: W81XWH-07-1-0390

TITLE: Targeting Signal Transducers and Activators of Transcription-3 (STAT3) as a Novel Strategy in Sensitizing Breast Cancer to EGFR-Targeted Therapy

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REPORT DATE: June 2009

TYPE OF REPORT: Annual

PREPARED FOR: U.S. Army Medical Research and Materiel Command  
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;  
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|--|--------------------------|---|---------------------------|--|
| 1. REPORT DATE<br>1 June 2009  | 2. REPORT TYPE<br>Annual | 3. DATES COVERED<br>1 June 2008 – 31 May 2009 |                           |  |
| 4. TITLE AND SUBTITLE<br><br>Targeting Signal Transducers and Activators of Transcription-3 (STAT3) as a Novel Strategy in Sensitizing Breast Cancer to EGFR-Targeted Therapy  |                          | 5a. CONTRACT NUMBER                           |                           |  |
|  |                          | 5b. GRANT NUMBER<br>W81XWH-07-1-0390          |                           |  |
|  |                          | 5c. PROGRAM ELEMENT NUMBER                    |                           |  |
| 6. AUTHOR(S)<br><br>Hui-Wen Lo, Ph.D.<br><br>E-Mail: huiwen.lo@duke.edu  |                          | 5d. PROJECT NUMBER                            |                           |  |
|  |                          | 5e. TASK NUMBER                               |                           |  |
|  |                          | 5f. WORK UNIT NUMBER                          |                           |  |
| 7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES)<br><br>Duke University<br>Durham, NC 27708  |                          | 8. PERFORMING ORGANIZATION REPORT NUMBER      |                           |  |
| 9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES)<br><br>U.S. Army Medical Research and Materiel Command<br>Fort Detrick, Maryland 21702-5012  |                          | 10. SPONSOR/MONITOR'S ACRONYM(S)              |                           |  |
|  |                          | 11. SPONSOR/MONITOR'S REPORT NUMBER(S)        |                           |  |
| 12. DISTRIBUTION / AVAILABILITY STATEMENT<br>Approved for Public Release; Distribution Unlimited   |                          |   |                           |  |
| 13. SUPPLEMENTARY NOTES  |                          |   |                           |  |
| 14. ABSTRACT<br>Our research effort in the past award year has resulted in several interesting findings that support the study hypothesis: deregulated EGFR and STAT3 pathways synergistically contribute to the malignant biology of breast cancer and that combined uses of anti-EGFR and anti-STAT3 treatments result in significantly increased breast cancer cell death compared to single agent treatments. First, we have created isogenic breast cancer cell lines to stably express modestly activated and highly activated STAT3, STATCA. Second, using these established isogenic breast cancer cell lines, we found that increased STAT3 activation rendered breast cancer cells resistant to EGFR-targeted therapy. Third, we showed that STAT3 expression knock-down sensitized EGFR-expressing breast cancer cells to anti-EGFR therapy. Finally, our in vitro results showed that combined used of a STAT3 small molecular weight inhibitor AG490 and a clinically used EGFR inhibitor Iressa synergistically targeted EGFR-expressing breast cancer cells to anti-EGFR therapy. |                          |   |                           |  |
| 15. SUBJECT TERMS<br>EGFR, STAT3, drug resistance, signal cross-talks  |                          |   |                           |  |
| 16. SECURITY CLASSIFICATION OF:<br><br>a. REPORT<br>U  |                          | 17. LIMITATION OF ABSTRACT<br>UU              | 18. NUMBER OF PAGES<br>10 | 19a. NAME OF RESPONSIBLE PERSON<br>USAMRMC |
| b. ABSTRACT<br>U   |                          |   |                           | 19b. TELEPHONE NUMBER (include area code)  |
| c. THIS PAGE<br>U  |                          |   |                           |  |

## Table of Contents

|  | Page     |
|--|----------|
| <b>Introduction.....</b>                 | <b>4</b> |
| <b>Body.....</b>                         | <b>4</b> |
| <b>Key Research Accomplishments.....</b> | <b>6</b> |
| <b>Reportable Outcomes.....</b>          | <b>6</b> |
| <b>Conclusion.....</b>                   | <b>7</b> |
| <b>References.....</b>                   | <b>7</b> |
| <b>Appendices.....</b>                   | <b>7</b> |

## INTRODUCTION

The proposal is built on the following observations from our studies and others. **(1)** Single use of EGFR-targeted therapy, gefitinib/Iressa (an EGFR tyrosine kinase inhibitor) and cetuximab (an EGFR blocking antibody), demonstrated a moderate therapeutic effect in breast cancer patients. **(2)** Both EGFR and STAT3 are oncoproteins and frequently over-expressed and/or constitutively activated in breast cancer (Lo et al. 2005b). We observed co-overexpression of both EGFR and p-STAT3 in 60% of EGFR-positive breast carcinomas (Lo et al. 2005a). **(3)** STAT3 is constitutively activated in 50% of the breast cancer and can be activated by EGFR and other growth factor- and cytokine pathways (Garcia et al. 1997; Yu and Jove 2004). **(4)** Iressa-treated breast cancer cells and clinical specimens displayed insufficient suppression of STAT3 activity despite with complete EGFR inhibition (Albanell et al. 2001; Lo et al. 2005a). **(5)** Although EGFR and STAT3 can mediate different downstream targets, our recent study indicated that they cross-talk at multiple levels (Lo et al. 2005a). Based on these rationales, we **hypothesize** that deregulated EGFR and STAT3 pathways synergistically contribute to the malignant biology of breast cancer and that combined uses of anti-EGFR and anti-STAT3 treatments result in significantly increased breast cancer cell death compared to single agent treatments. The hypothesis is being tested by the following three Specific Aims.

**AIM 1:** To determine whether increased STAT3 activity confers resistance to anti-EGFR therapies in EGFR-expressing breast cancer cells.

**AIM 2:** To investigate whether suppression of STAT3 expression/activity sensitizes EGFR/p-STAT3-expressing breast cancer cells to anti-EGFR therapies.

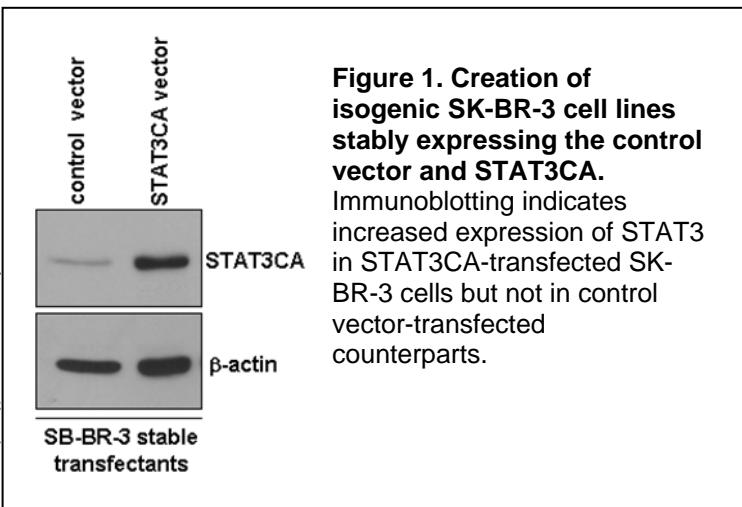
**AIM 3:** To determine the therapeutic effects of combined use of anti-EGFR and anti-STAT3 treatments in a mammary tumor-bearing animal model.

The outcome from this proposal is likely to help us achieve the long-term goal of the study, which is to better understand the malignant biology of breast cancer including those with de-regulated EGFR pathway and to provide rationales for more effective therapies for women with breast cancer.

## BODY

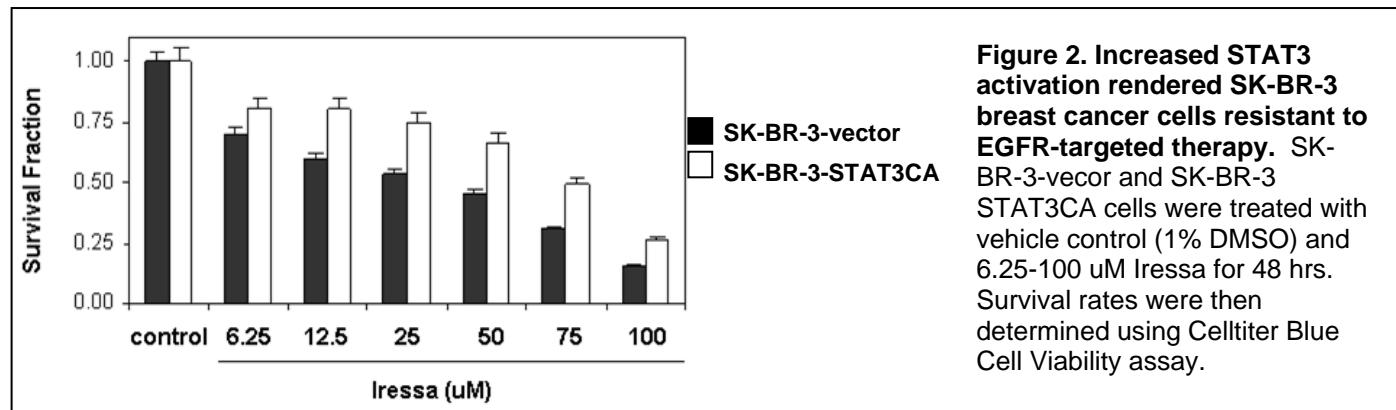
**Creation of isogenic breast cancer cells that stably express modestly and highly activated STAT3 (Task 1-b).** To determine whether increased STAT3 expression/activity confers resistance to anti-EGFR therapy in EGFR-expressing breast cancer cells, we first generated isogenic SK-BR-3 breast cancer cell lines to express the control vector or constitutively activated STAT3, STAT3CA. SK-BR-3 cells express modest levels of activated STAT3 and are relatively sensitive to Iressa, and are thus ideal for the proposed study. It is worthwhile to mention that STAT3CA is a modified variant of STAT3 that contains substitution of two Cys residues within the SH2 domain and dimerizes spontaneously, binds to DNA, activates transcription, and transforms normal fibroblasts (Bromberg et al. 1999). More recently, STAT3CA has been linked to tumorigenesis of prostate (Qin et al. 2008) and skin (Pedranzini et al. 2004; Chan et al. 2004). STAT3CA also transforms mouse bone marrow cells into highly aggressive T cell leukemia in mice (Ecker et al. 2009).

As shown by the immunoblotting in **Figure 1**, we have successfully established two isogenic SK-BR-3 breast cancer stable transfectants to express the control vector and STAT3CA vector. In these studies, we transfected SK-BR-3 cells with corresponding vectors and subjected them to G418 treatments for one month to allow for stable selection. Stable clones were then examined for STAT3CA expression using immunoblotting. Together, these results indicate that these isogenic cells are ideal for examining the effects of increased STAT3 activity on the resistance of breast cancer cells to EGFR-targeted therapy (Task 1-c).

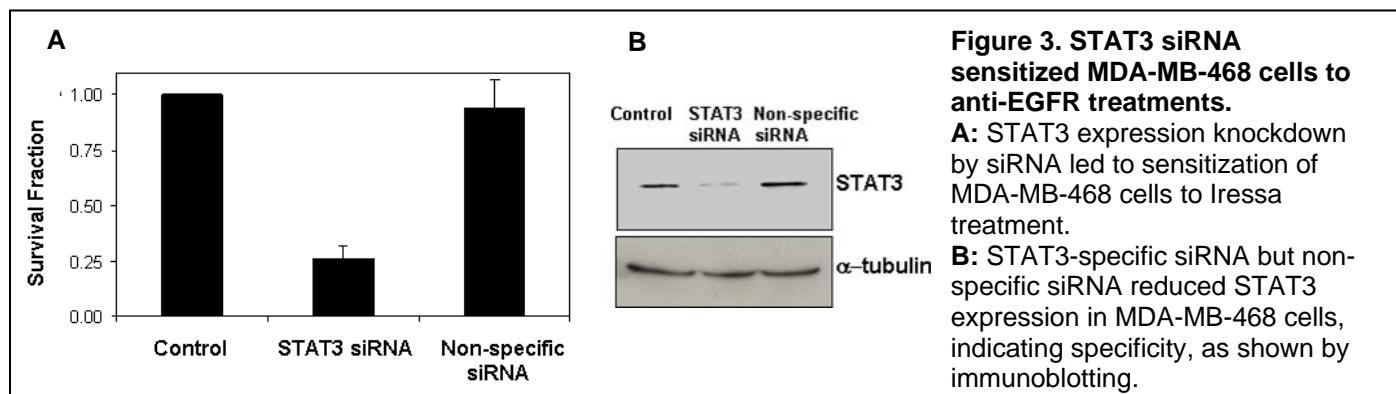


### Increased STAT3 activation rendered breast cancer cells resistant to EGFR-targeted therapy

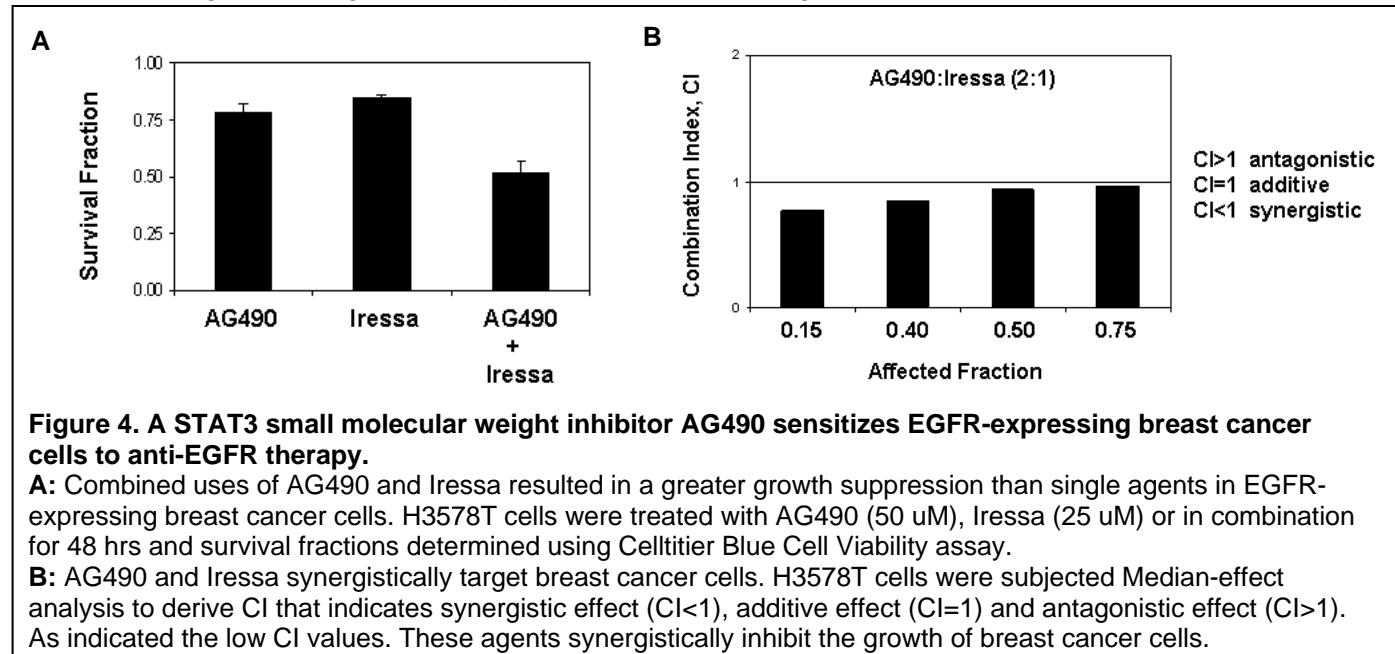
**(Task 1-c).** Using the isogenic SK-BR-3 cell lines, we examined their response to various concentrations of Iressa following 48 hr treatments. Cell survival was determined using Celltiter Blue Cell Viability assay kit (Promega), a fluorescent method that is based on the ability of living cells to convert a redox dye (resazurin) into a fluorescent end product, resorufin, as we previously described (Lo et al., 2008a,b). As shown by the results in **Figure 2**, SK-BR-3-STAT3CA cells were more resistant to Iressa-mediated growth suppression compared to SK-BR-3-vector cells. These results indicate that increased STAT3 activity rendered breast cancer cells more resistant to anti-EGFR agent, Iressa.



**STAT3 expression knock-down sensitizes EGFR-expressing breast cancer cells to anti-EGFR therapy (Task 2-a).** In the 2008-progress report, we reported that suppressing STAT3 activity using dominant-negative STAT3 mutant (STAT3-DN) led to increased sensitivity of breast cancer cells to EGFR-targeted therapy. Here, we further suppressed STAT3 expression using STAT3-specific siRNA to determine the effects of this on sensitivity of breast cancer cells to Iressa. In these studies, we used MDA-MB-468 breast cancer cells that are known to express high levels of both EGFR and p-STAT3 (Y705) and resistant to EGFR-targeted agents, as we previously reported (Lo et al. 2005a). These cells were untransfected or transfected with STAT3-specific siRNA or non-specific siRNA (Dharmacon). Forty-eight hrs following transfactions, the cells were subjected to cell viability assay, as we described earlier. As shown by the results in **Figure 3A**, STAT3 expression down-regulation sensitized MDA-MB-468 cells to Iressa treatment. Non-specific siRNA did not affect cell survival indicating assay specificity. As expected, STAT3 siRNA specifically and effectively reduced STAT3 protein expressing, as indicated by immunoblotting (**Figure 3B**). Collectively, the results from the STAT3 siRNA experiments indicate that targeting STAT3 sensitizes EGFR-expressing breast cancer cells to EGFR-targeted therapy *in vitro*.



**A STAT3 small molecular weight inhibitor AG490 sensitizes EGFR-expressing breast cancer cells to anti-EGFR therapy (Task 2-b).** To further investigate Specific Aim 2, we determined the extent to which STAT3 inhibition sensitizes breast cancer cells to EGFR-targeted therapy using AG490, a small molecular weight inhibitor that targets STAT3-activating kinase, JAK2. Interestingly, we found breast cancer cells to respond to AG490+Iressa combination better than AG490 alone and Iressa alone (**Figure 4A**). Using the Median-effect analysis (Chou and Talalay 1984) that computes combination index (CI), we determined whether AG490 and Iressa targeted breast cancer cells synergistically, as we previously described (Lo et al. 2008). As indicated by the low CI values, we found combination of STAT3 and EGFR inhibitors to be synergistic in targeting breast cancer cells (**Figure 4B**). Collectively, the results from STAT3-DN (2008-report), STAT3 siRNA experiments (Figure 3) and STAT3 inhibitor study (Figure 4) indicate that targeting STAT3 sensitizes EGFR-expressing breast cancer cells to EGFR-targeted therapy *in vitro*. These *in vitro* results also strongly justify the *in vivo* studies proposed in Specific Aim 3 to examine the efficacy of the combination regimen using an orthotopic breast cancer xenograft model.



**Figure 4. A STAT3 small molecular weight inhibitor AG490 sensitizes EGFR-expressing breast cancer cells to anti-EGFR therapy.**

**A:** Combined uses of AG490 and Iressa resulted in a greater growth suppression than single agents in EGFR-expressing breast cancer cells. H3578T cells were treated with AG490 (50  $\mu$ M), Iressa (25  $\mu$ M) or in combination for 48 hrs and survival fractions determined using Celltiter Blue Cell Viability assay.

**B:** AG490 and Iressa synergistically target breast cancer cells. H3578T cells were subjected Median-effect analysis to derive CI that indicates synergistic effect (CI < 1), additive effect (CI = 1) and antagonistic effect (CI > 1). As indicated the low CI values. These agents synergistically inhibit the growth of breast cancer cells.

## KEY RESEARCH ACCOMPLISHMENTS

- Task 1-b: Establishment of isogenic breast cancer cells that stably express modestly and highly activated STAT3.
- Task 1-c: Increased STAT3 activation rendered breast cancer cells resistant to EGFR-targeted therapy.
- Task 2-a: STAT3 expression knock-down sensitizes EGFR-expressing breast cancer cells to anti-EGFR therapy.
- Task 2-b: A STAT3 small molecular weight inhibitor AG490 sensitizes EGFR-expressing breast cancer cells to anti-EGFR therapy.

## REPORTABLE OUTCOMES

### Funding applied based on work supported by this award

Research Scholar Grant Lo (PI)  
 American Cancer Society  
 De-regulated EGFR and STAT3 pathways in breast cancer EMT and intravasation

## CONCLUSION

Our research effort in the past award year has resulted in several interesting findings that support the study hypothesis: deregulated EGFR and STAT3 pathways synergistically contribute to the malignant biology of breast cancer and that combined uses of anti-EGFR and anti-STAT3 treatments result in significantly increased breast cancer cell death compared to single agent treatments. First, using established isogenic breast cancer cells that stably express modestly and highly activated STAT3, we found Increased STAT3 activation rendered breast cancer cells resistant to EGFR-targeted therapy. Second, we found that STAT3 expression knock-down sensitizes EGFR-expressing breast cancer cells to anti-EGFR therapy. Finally, our results showed that combined used of a STAT3 small molecular weight inhibitor AG490 and a clinically used EGFR inhibitor synergistically target EGFR-expressing breast cancer cells to anti-EGFR therapy. Together, these promising results prompt us to further explore the role of STAT3 activation in the resistance of human breast cancer cells to anti-EGFR therapy in animals in the next award year.

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Yu, H., and R. Jove. 2004. The STATs of cancer--new molecular targets come of age. *Nat Rev Cancer* 4 (2):97-105.

## APPENDIX

### Updated Biosketch

## BIOGRAPHICAL SKETCH

| NAME<br><b>LO, HUI-WEN, Ph.D.</b>  | POSITION TITLE<br><b>Assistant Professor</b> |         |                                    |
|--|--|---------|------------------------------------|
| eRA COMMONS USER NAME<br>HUIWENLO  |  |         |                                    |
| EDUCATION/TRAINING (Begin with baccalaureate or other initial professional education, such as nursing, and include postdoctoral training.) |  |         |                                    |
| INSTITUTION AND LOCATION   | DEGREE<br>(if applicable)                    | YEAR(s) | FIELD OF STUDY                     |
| Chung-Shan Medical & Dental College  | B.S.   | 1986    | Nutrition                          |
| The University of Texas at Austin  | M.A.   | 1990    | Nutritional Sciences               |
| The University of Texas-Health Science Center at Houston   | M.S.   | 1994    | Biomedical Sciences                |
| The University of Texas-Health Science Center at Houston   | Ph.D.  | 2002    | Biochemistry and Molecular Biology |
| MD Anderson Cancer Center  | POSTDOC                                      | 2004    | Cancer Research                    |

### **A: POSITIONS AND HONORS**

#### **Positions and Employment**

|              |   |
|--------------|---|
| 2006-present | Assistant Professor (tenure-track), Department of Surgery, Duke University School of Medicine |
| 2005-2006    | Instructor, University of Texas M.D. Anderson Cancer Center                                   |
| 2002-2004    | Postdoctoral Fellow, University of Texas M.D. Anderson Cancer Center                          |
| 1998-2000    | Research Investigator, University of Texas M.D. Anderson Cancer Center                        |
| 1993-1998    | Senior Research Assistant, University of Texas M.D. Anderson Cancer Center                    |

#### **Awards and Honors (selected)**

|              |   |
|--------------|---|
| 2008         | Outstanding Alumni Award, Chung-Shan Medical University, Taichung, Taiwan                                       |
| 2008         | Career Development Award, Duke Brain Cancer SPORE, NCI  |
| 2007-present | Idea Award, Department of Defense   |
| 2006-present | The Howard Temin Award, NCI   |
| 2005-2006    | American Cancer Society Postdoctoral Fellowship   |
| 2004         | AFLAC Scholar-in-Training Award, American Association for Cancer Research                                       |
| 2004         | Trainee Excellence Award, M.D. Anderson Cancer Center   |
| 2002         | WICR Brigid G. Leventhal Scholar Award, American Association for Cancer Research-Women in Cancer Research       |
| 2001         | John P. McGovern Award, Graduate School of Biomedical Sciences<br>The University of Texas-Health Science Center |

#### **Professional Memberships**

|              |   |
|--------------|---|
| 2007-present | Member, Duke Comprehensive Cancer Center                                  |
| 2006-present | Member, Society of Neuro-oncology   |
| 1997-present | Member, American Association for Cancer Research                          |
| 1997-present | Member, Women in Cancer Research-American Association for Cancer Research |

### **B. SELECTED PEER-REVIEWED PUBLICATIONS (FROM 24 PUBLICATIONS)**

1. **Lo, H.-W.** and Ali-Osman, F. Genomic cloning of hGSTP1\*C, an allelic human pi class glutathione S-transferase gene variant and functional characterization of its retinoic acid response elements. *Journal of Biological Chemistry* 272:32743-32749, 1997.

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## C. RESEARCH SUPPORT

### ACTIVE:

**The Howard Temin Award (5K01-CA118423-03)** (Lo, PI)

9/25/2006-7/31/2011

National Cancer Institute

Nuclear EGFR Signaling Network in Human Cancers

The goal of this application is to understand the biological role of the nuclear EGFR signaling pathway in human cancers. Aim 1: Characterize the transcriptional co-regulation of the iNOS gene by nuclear interaction of EGFR and STAT3 and determine its role in tumor survival. Aim 2: Characterize interaction of nuclear EGFR with c-jun and determine its effect on TWIST gene activation and TWIST-mediated breast cancer progression. Aim 3: determine the role of nuclear EGFR and underlying mechanisms in chemo-resistance.

**Idea Award (W81XWH-07-1-0390)**

(Lo, PI)

6/1/2007-6/30/2010

Breast Cancer Research Program

Department of Defense

Targeting signal transducer and activator of transcription 3 as a novel strategy in sensitizing breast cancer to anti-EGFR therapy

The goal of this project is to determine the role of STAT3 activation in the resistance of breast cancer to EGFR-targeted therapy. AIM 1: To determine whether increased STAT3 expression/activity confers resistance to anti-EGFR therapy in EGFR-expressing breast cancer cells. AIM 2: To investigate whether suppression of STAT3 expression/activity sensitizes EGFR-expressing breast cancer cells to anti-EGFR therapy. AIM 3: To determine the therapeutic effects of combined use of anti-EGFR and anti-STAT3 treatments in a mammary tumor-bearing animal model.